

NON-INVASIVE OBJECTIVE ASSESSMENT OF DIABETIC FOOT ULCER  
HEALING WITH BLOOD PERFUSION AND TISSUE OXYGENATION

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To my beloved parents, thank you.



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## ABSTRACT

The high prevalence of the world's population diagnosed with diabetes mellitus, with a significant number suffering from diabetic foot ulcer (DFU), has always been a global concern. Although modern medical technologies are available to assist medical physician in wound diagnosis, the practicability of these techniques have yet to be critically assessed. Only a limited number of biomedical researches have examined the correlation between blood flow and tissue oxygenation in diabetic wounds. This thesis is a pioneering work that aims to develop a reliable diagnostic tool to address the tremendous need for coordinated and efficient DFU management via prediction of relative blood perfusion ( $\tau$ ) and transcutaneous oxygen saturation ( $S_tO_2$ ). The laser speckle integrated multispectral imaging system is an optical, non-invasive system that is able to provide quantitative and visual information of blood perfusion and tissue oxygen in diabetic ulcer. The estimation of blood perfusion is based on speckle contrast analysis of blood flow whereas the estimation of  $S_tO_2$  parameter is by means of fitting the Extended Modified Lambert Beer model to the collected attenuation data. This system incorporates the use of a 650 nm low power laser diode for *in-vivo* assessment of blood perfusion and wavelength in the visible range of 530–570 nm to predict tissue oxygenation using *priori* information of hemoglobin's coefficients. A study was conducted on DFU patients recruited from Hospital Sultanah Nora Ismail. The results from this research revealed a slightly higher mean blood perfusion and tissue oxygen level in positive healing wounds than in impaired healing wounds, despite data indicating no statistical significance between these two groups (  $\rho=0.909$  and  $\rho=0.512$  for  $\tau$  and  $S_tO_2$  data, respectively). This research concludes that a mean blood perfusion index of  $1.5 (\times 10^3)$  and percent  $S_tO_2$  of at least 70 % are vital during the proliferative phase to ensure progressive healing. The strategies explored in this work can provide quantitative information of changes in blood perfusion with tissue revascularization to evaluate the outcomes of skin grafting procedures in positive and impaired healing diabetic wounds.



## ABSTRAK

Kelaziman populasi dunia yang didiagnosis dengan *diabetes mellitus*, dengan sejumlah besar menderita ulser kaki diabetes (DFU), menjadi kebimbangan global sejak beberapa tahun lalu. Walaupun terdapat teknologi perubatan moden untuk membantu doktor perubatan dalam diagnosis luka, kebolehlaksanaan teknik ini belum dinilai secara kritikal. Hanya sebilangan kecil penyelidikan bioperubatan yang mengkaji kaitan antara aliran darah dan oksigenasi tisu pada luka diabetes. Tesis ini merupakan usaha perintis yang bertujuan untuk membangunkan alat diagnostik bagi mengatasi keperluan besar pengurusan DFU yang selaras dan efisien melalui pengukuran perfusi darah relatif ( $\tau$ ) dan ketepuan oksigen transkutan ( $S_tO_2$ ). Sistem pengimejan bersepadu bintik laser dan multispektral adalah sistem optik tidak invasif yang dapat menyediakan maklumat kuantitatif dan visual oksigen tisu dan peredaran darah dalam ulser diabetik. Anggaran parameter  $S_tO_2$  adalah dengan memadankan model *Extended Modified Lambert Beer* kepada data pelemahan yang dikumpul manakala anggaran perfusi darah berdasarkan analisis bintik kontras aliran darah. Sistem ini menggabungkan penggunaan diod laser kuasa rendah 650 nm untuk taksiran perfusi darah *in-vivo* serta panjang gelombang nampak dalam rangkaian 530 – 570 nm untuk anggaran pengoksigenan tisu menggunakan maklumat *priori* pekali hemoglobin. Kajian dijalankan terhadap pesakit DFU yang direkrut dari Hospital Sultanah Nora Ismail. Hasil kajian ini mendedahkan tahap perfusi darah dan oksigen tisu yang lebih tinggi dalam luka penyembuhan positif berbanding luka penyembuhan terencat walaupun data menunjukkan tiada keertian statistik antara kedua-dua kumpulan ( $\rho = 0.909$  dan  $\rho = 0.512$  masing-masing untuk  $\tau$  dan  $S_tO_2$ ). Kajian ini menyimpulkan bahawa min indeks perfusi darah  $1.5 (\times 10^3)$  dan peratus  $S_tO_2$  sekurang-kurangnya 70 % adalah penting semasa fasa proliferasif untuk menjamin penyembuhan progresif. Strategi yang diterokai dalam kerja ini dapat memberi maklumat kuantitatif perubahan dalam perfusi darah dengan vaskularisasi tisu dalam penyembuhan luka diabetik positif dan terencat.

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$\tau = 1.3 (\times 10^3)$  and  $S_tO_2 = 75\%$ . The red bar

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## LIST OF SYMBOLS AND ABBREVIATIONS

$A$	–	Light attenuation
$C$	–	Concentration of absorber, mol L <sup>-1</sup>
CCD	–	Charge-coupled detector
$d$	–	Light pathlength, mm
$D$	–	Aperture diameter, mm
DFU	–	Diabetic foot ulcer
EMLB	–	Extended Modified Lambert Beer
$f$	–	Frequency, hertz
$f_L$	–	Focal length, mm
$G_0$	–	Light attenuation offset
$G_1$	–	Absorption dependent light attenuation
Hb	–	Deoxygenated hemoglobin
HbO <sub>2</sub>	–	Oxyhemoglobin
HeNe	–	Helium-Neon
$I_0$	–	Transmitted light intensity
$I$	–	Detected light intensity
$\langle I \rangle$	–	Mean light intensity
$k$	–	Speckle contrast
LASCA	–	Laser speckle contrast analysis
NIR	–	Near-infrared
NMRR	–	National Medical Research Register
$S$	–	Speckle size
S <sub>a</sub> O <sub>2</sub>	–	Arterial blood oxygen saturation, %
S <sub>t</sub> O <sub>2</sub>	–	Transcutaneous oxygen saturation, %
SNR	–	Signal to noise ratio, dB
$T$	–	Light transmittance
$T_H$	–	Total absorbers concentration, mol L <sup>-1</sup>

$T_s$	–	Exposure time, second
$v$	–	Velocity, $\text{ms}^{-1}$
$\varepsilon$	–	Molar extinction coefficient, $\text{mm}^{-1} \text{M}^{-1}$
$\Delta\varepsilon$	–	Absorptivity difference, $\text{mm}^{-1} \text{M}^{-1}$
$\lambda$	–	Wavelength, nm
$\mu_a$	–	Absorption coefficient, $\text{mm}^{-1}$
$\mu_s$	–	Scattering coefficient, $\text{mm}^{-1}$
$\rho$	–	Probability value
$\rho\text{O}_2$	–	Partial pressure of oxygen, mmHg
$\sigma$	–	Standard deviation
$\tau$	–	Correlation time
$\phi$	–	Diameter, mm



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## **CHAPTER 1**

### **INTRODUCTION**

#### **1.1 Background of the study**

A retrospective study by the World Health Organization (WHO) based on data sources collected from 221 countries and territories had estimated at least 451 million adults aged 18–99 years old worldwide diagnosed with diabetes mellitus as of the year 2017 [1]. This figure was expected to project up to 693 million by the year 2045. A majority of diabetes cases were identified among those living in low- and middle- income countries, with Western Pacific region having the highest prevalence of total global diabetes population. China was quoted as having the highest number of population with diabetes with 98.4 million adults diagnosed, followed by India with 65.1 million [2]. Likewise in Malaysia, a statistical study conducted by the National Health and Morbidity Surveys (NHMS) in the year 2011 reported at least 15.2 % of adults aged 18 years old and above suffered from diabetes mellitus, which accounted for up to 2.6 million of the population [3]. The prevalence of this disease was expected to escalate to 21.6 % by the year 2020. Although there was no significant difference in prevalence between gender, a large variation was observed between the three main ethnic groups in Malaysia with the highest prevalent among Indians at 24.9 %, followed by Malays at 16.9 % and finally, Chinese at 13.8 % [4]. Specifically, a retrospective study by Raja [5] stated that diabetic foot ulcer (DFU), a microvascular complication of diabetes mellitus, constituted 15 % of all diabetes cases. Of this figure, approximately 20 % of DFU cases are the results of peripheral and autonomic neuropathy as well as diabetic angiopathy, which, if left untreated can lead to foot infection and amputation of the lower extremity. Socio-economic status, living environment and eating habits are

among the factors linked to the radical rise in diabetes cases worldwide [6]. The global health expenditure for diabetes patients in 2017 was estimated to be USD 850 billion, which was twice the expenditure of patients without diabetes [1]. Moreover, the number of deaths attributed to diabetes in the same year was reported to be approximately five million. The occurrence of this disease had been deemed to be increasingly alarming as it had left deleterious effects on impacted patients and their families both emotionally and financially.

Thus far, biomedical researchers worldwide have proposed various techniques to overcome the tradeoffs of standard clinical practices. A reliable and non-invasive technique for quantitative measurement of DFU healing progress has been a subject widely researched in recent years. The conventional method of bedside clinical assessment by means of subjective visual assessment, although can be performed immediately at no cost, has otherwise turned out to be unreliable and inaccurate in terms of diagnosis [7]. Over the years, several techniques to predict wound healing status have been introduced. These techniques, for example, range from punch biopsy method for bacterial load analysis, which is not only invasive in nature but also time-consuming [8], to non-invasive optical imaging approach such as Raman spectroscopy technique that involves tedious tabulation works [9]. Moreover, the commonly practiced laser Doppler flowmetry is only able to provide information on blood flow perfusion measurement in tissue [10]. Therefore, the limitations of the use of these techniques are critically assessed [11].

Meanwhile, modern clinical practice of diagnosing and monitoring the progression of diseases involves the use of medical imaging techniques. Among the commonly used modalities at present are magnetic resonance imaging (MRI), computerized tomography (CT) scan, positron emission tomography (PET) scan and single photo emission computed tomography (SPECT) scan [12-14]. These cutting edge imaging techniques are proven to be highly beneficial in detecting any abnormalities in the tissue structure based on two-dimensional (2D) or three-dimensional (3D) images that produce very detailed anatomic information of the scanned body. However, these machines are costly in price and require the use of contrast agents or radioactive tracers to be administered into the patient's body, hence, limiting the application of these approaches on patients with critical illnesses since the use of contrast agents has side effects likely not recommended for pregnant women [15, 16].

Early prognosis and evaluation of DFU recovery progress can be performed using quantitative information deduced from changes in skin microcirculatory activities namely tissue oxygenation and blood perfusion level at different stages of wound healing. Sufficient blood perfusion and oxygen supply to wound regions are likely to guarantee positive healing progress, hence the requirement of a systematic prognosis system to provide clinical information of diabetic ulcer microcirculatory status. Proper wound care management is not only vital to achieve full recovery of the anatomical functionality; it also contributes to the reduction in medical cost in the long period. This is possible given that medical practitioners are provided with adequate information as a guideline to identify detrimental factors leading to delay of healing in DFU and immediate actions are undertaken to overcome these problems.

Awareness of this matter has led to the rise in biomedical research to come up with an applicable diagnosis tool that can provide detailed *in-vivo* information of wound based on the optical properties of scattering and absorbing media in the biological skin tissue. This is viable using a spectroscopic and spatially resolved speckle interference pattern obtained from a speckle integrated multispectral imaging system. Laser speckle contrast imaging is a technique which has advanced over the years, with its application ranging from imaging of multimodal skin layers, cerebral cortex, and retina using light of wavelength 633 nm [17, 18]. The fundamental basis of this concept is based on the interference pattern of light reflected or scattered from the illuminating surface, which in return produces a granular effect known as the laser speckle. The movement of the speckle pattern is concurrent to the movement of an object, prompting the idea of the most significant application of speckle fluctuation initiated by blood flow velocity during the mid-1970s [19].

This work sets out to employ laser speckle integrated multispectral imaging for concurrent measurement of blood perfusion and oxygen saturation mapping of foot ulcers. The best guess of these required values is via a heuristically developed fitting model predicted based on light absorption properties and time-integrated speckle pattern produced by laser light interference between the light source and refraction in the medium. To further verify the effectiveness of this method, efforts are undertaken to collaborate with a hospital where a group of DFU patients was recruited as volunteers to support this study with solid scientific and clinical data. This study presumes the proposed quantitative values of wound healing parameters and the developed strategies can assist surgeons in determining the uncertainty of

wound closure through proper classification of chronic diabetic foot ulcers healing status.

## 1.2 Problem statement

There is a tremendous need for coordinated and efficient wound healing management in DFU patients nationwide. The predominance of DFU among patients suffering from diabetes has led to limitations in daily activities as well as reduced living quality. In some cases, chronic DFU can cause morbidity and lower limb amputation that results in permanent disability and also contributes to emotional depression. The call for proper management and assessment of diabetic wound is highly sought after with the increase in the use of healthcare centers and hospitals. As the economy continues to decline and the cost of medical fee increases, the need for a more cost-effective and systematic wound management solution expands throughout the nation [20]. Undoubtedly, there are many medical options for accurate diagnosis of DFU, but these options lack in terms of suitability and can be costly in price [15, 16]. Numerous methods have been proposed in the biomedical field of study to improve the prediction accuracy of wound healing progress, taking into account aspects such as time, cost and consistency [7, 21, 22]. However, these works were unable to concurrently image blood flow and tissue oxygenation status during the different phases of DFU healing progress. As such, a non-invasive and detailed approach to study the optical properties of the biological skin tissue is highly sought after. The laser speckle contrast technique has been proven capable in measuring cutaneous blood flow whereas multispectral imaging is a non-invasive approach to acquire optical properties of biological tissue [23-25]. No work, has, thus far, combined the two technologies. In addition, assessment of wound should include a complete understanding of wound pathology, environmental factors that affect healing and intense study on tissue remodeling. These environmental factors, which include microenvironment factors of that within or surrounding the wound such as blood flow, moisture, nutrient availability and bioburden factors, e.g. biofilm concentration are among the factors accounted for when ensuring a conducive environment for healing [26]. This allows a strategic treatment to be assigned accordingly to each patient bearing these factors to ensure a progressive healing. A strong correlation

## REFERENCES

1. Cho, N., Shaw, J., Karuranga, S., Huang, Y., da Rocha Fernandes, J., Ohlrogge, A., and Malanda, B. IDF Diabetes Atlas: Global Estimates of Diabetes Prevalence for 2017 and Projections for 2045. *Diabetes Research and Clinical Practice*. 2018. 138: 271-281.
2. Guariguata, L., Whiting, D.R., Hambleton, I., Beagley, J., Linnenkamp, U., and Shaw, J.E. Global Estimates Of Diabetes Prevalence for 2013 and Projections for 2035. *Diabetes Research and Clinical Practice*. 2014. 103(2): 137-149.
3. Feisul, M., and Azmi, S. *National Diabetes Registry Report, Volume 1, 2009–2012*. Kuala Lumpur: Ministry of Health, Malaysia. 2013.
4. Institute for Public Health. *National Health and Morbidity Survey 2011 (NHMS 2011)*. Malaysia Ministry of Health. 2011.
5. Raja, N.S. Microbiology of Diabetic Foot Infections in a Teaching Hospital in Malaysia: A Retrospective Study of 194 Cases. *Journal of Microbiology Immunology and Infection*. 2007. 40(1): 39.
6. Purwaningsih, T., and Machmud, B. Influence of Overweight and Obesity on the Diabetes in the World on Adult People Using Spatial Regression. *International Journal of Advances in Intelligent Informatics*. 2016. 2(3): 149-156.
7. Maisel, W.H., and Lewis, R.J. Noninvasive Measurement Of Carboxyhemoglobin: How Accurate is Accurate Enough. *Annals Of Emergency Medicine*. 2010. 56(4): 389-391.
8. Xu, L., McLennan, S.V., Lo, L., Natfaji, A., Bolton, T., Liu, Y., Twigg, S.M., and Yue, D.K. Bacterial Load Predicts Healing Rate in Neuropathic Diabetic Foot Ulcers. *Diabetes Care*. 2007. 30(2): 378-380.
9. Mao, X. *Evaluation of Chronic Wounds by Raman Spectroscopy and Image Processing*. Ph.D. Thesis, Drexel University; 2012.

10. Humeau-Heurtier, A., Guerreschi, E., Abraham, P., and Mahé, G. Relevance of Laser Doppler and Laser Speckle Techniques for Assessing Vascular Function: State of the Art and Future Trends. *IEEE Transactions On Biomedical Engineering*. 2013. 60(3): 659-666.
11. Barry, E., Roberts, S., Oke, J., Vijayaraghavan, S., Normansell, R., and Greenhalgh, T. Efficacy and Effectiveness of Screen and Treat Policies in Prevention of Type 2 Diabetes: Systematic Review and Meta-Analysis of Screening Tests and Interventions. *bmj*. 2017. 356: i6538.
12. Long, Z., Hanson, D.P., Mullan, B.P., Hunt, C.H., Holmes III, D.R., Brinkmann, B.H., and O'Connor, M.K. Analysis of Brain Spect Images Coregistered with Mri in Patients with Epilepsy: Comparison of Three Methods. *Journal of Neuroimaging*. 2018. 28(3): 307-312.
13. Seeram, E. *Computed Tomography: Physical Principles, Clinical Applications, and Quality Control*. Elsevier Health Sciences. 2015.
14. Flotats, A., Knuuti, J., Gutberlet, M., Marcassa, C., Bengel, F.M., Kaufmann, P.A., Rees, M.R., and Hesse, B. Hybrid Cardiac Imaging: SPECT/CT and PET/CT. A Joint Position Statement by the European Association of Nuclear Medicine (EANM), the European Society of Cardiac Radiology (ESCR) and the European Council of Nuclear Cardiology (ECNC). *European Journal Of Nuclear Medicine And Molecular Imaging*. 2011. 38(1): pp. 201-212.
15. Padole, A., Ali Khawaja, R.D., Kalra, M.K., and Singh, S.: CT Radiation Dose and Iterative Reconstruction Techniques'. *American Journal Of Roentgenology*. 2015. 204(4): W384-W392.
16. Desiderio, M.C., Lundbye, J., Baker, W.L., Farrell, M.B., and Heller, G. Contemporary Patient Radiation Exposure from PET Versus Spect MPI. *Journal of the American College of Cardiology*. 2018. 71(11): A1489.
17. Dunn, A.K. Laser Speckle Contrast Imaging of Cerebral Blood Flow. *Annals of Biomedical Engineering*. 2012. 40(2): 367-377.
18. Bliedtner, K., Seifert, E., Stockmann, L., Effe, L., and Brinkmann, R. Towards Real Time Speckle Controlled Retinal Photocoagulation. In: *Towards Real Time Speckle Controlled Retinal Photocoagulation*. International Society for Optics and Photonics. pp. 96931A; 2016.
19. Briers, D., Duncan, D.D., Hirst, E., Kirkpatrick, S.J., Larsson, M., Steenbergen, W., Stromberg, T., and Thompson, O.B. Laser Speckle Contrast



- Imaging: Theoretical and Practical Limitations. *Journal of Biomedical Optics*. 2013. 18(6): 066018-066018.
20. Bakker, K., Apelqvist, J., Lipsky, B., Van Netten, J., Schaper, N., and International Working Group on the Diabetic Foot (IWGDF). The 2015 IWGDF Guidance Documents on Prevention and Management of Foot Problems in Diabetes: Development of an Evidence-Based Global Consensus. *Diabetes/Metabolism Research and Reviews*. 2016. 32: 2-6.
  21. Richards, L.M., Kazmi, S.S., Davis, J.L., Olin, K.E., and Dunn, A.K. Low-Cost Laser Speckle Contrast Imaging of Blood Flow Using a Webcam. *Biomedical Optics Express*. 2013. 4(10): 2269-2283.
  22. Ravichandran, P., and Chitti, S.P. Antimicrobial Dressing for Diabetic Foot Ulcer Colonized with MRSA. *OnLine Journal of Biological Sciences*. 2015. 15 (4): 282.
  23. Wang, J., Wang, Y., Li, B., Feng, D., Lu, J., Luo, Q., and Li, P. Dual-Wavelength Laser Speckle Imaging to Simultaneously Access Blood Flow, Blood Volume, and Oxygenation Using a Color CCD Camera. *Optics Letters*. 2013. 38(18): 3690-3692.
  24. Huong, A., Philimon, S., and Ngu, X. Multispectral Imaging of Acute Wound Tissue Oxygenation. *Journal of Innovative Optical Health Sciences*. 2017. 10(03): 1750004.
  25. Shapey, J., Xie, Y., Nabavi, E., Bradford, R., Saeed, S.R., Ourselin, S., and Vercauteren, T. Intraoperative Multispectral and Hyperspectral Label-Free Imaging: A Systematic Review of in Vivo Clinical Studies. *Journal Of Biophotonics*. 2019. e201800455.
  26. Kruse, C.R., Nuutila, K., Lee, C.C., Kiwanuka, E., Singh, M., Caterson, E.J., Eriksson, E., and Sørensen, J.A. The External Microenvironment of Healing Skin Wounds. *Wound Repair and Regeneration*. 2015. 23(4): 456-464.
  27. Lindley, L.E., Stojadinovic, O., Pastar, I., and Tomic-Canic, M. Biology and Biomarkers for Wound Healing. *Plastic and Reconstructive Surgery*. 2016. 138(3): 18S.
  28. Iyer, A., Jeyalatha, S., and Sumbaly, R. Diagnosis of Diabetes Using Classification Mining Techniques. *arXiv preprint arXiv:1502.03774*. 2015.
  29. Asmat, U., Abad, K., and Ismail, K. Diabetes Mellitus and Oxidative Stress- A Concise Review. *Saudi Pharmaceutical Journal*. 2016. 24(5): 547-553.

30. Han, S., Middleton, P., Shepherd, E., Van Ryswyk, E., and Crowther, C.A. Different Types of Dietary Advice for Women with Gestational Diabetes Mellitus. *Cochrane Database of Systematic Reviews*. 2017. 2: CD009275.
31. Ramsey, S.D., Newton, K., Blough, D., McCulloch, D.K., Sandhu, N., Reiber, G.E., and Wagner, E.H. Incidence, Outcomes, and Cost of Foot Ulcers in Patients with Diabetes. *Diabetes Care*. 1999. 22(3): 382-387.
32. Wagner Jr, F.W. The Dysvascular Foot: A System for Diagnosis and Treatment. *Foot & Ankle*. 1981. 2(2): 64-122.
33. Korkmaz, M., Erdoğan, Y., Balci, M., Amanvermez Şenarslan, D., and Yılmaz, N. Preoperative Medical Treatment in Patients Undergoing Diabetic foot Surgery with a Wagner Grade-3 or Higher Ulcer: A Retrospective Analysis of 52 Patients. *Diabetic Foot & Ankle*. 2012. 3(1): 18838.
34. Stang, D., Young, M., and is Specialist, D.S. Selection and Application of a Diabetic Foot Ulcer Classification System in Scotland: Part 2. *The Diabetic Foot Journal*. 2018. 21(2): 100-106.
35. Lavery, L.A., Armstrong, D.G., and Harkless, L.B. Classification of Diabetic Foot Wounds. *The Journal of Foot and Ankle Surgery*. 1996. 35(6): 528-531.
36. Everett, E., and Mathioudakis, N.: Update on Management of Diabetic Foot Ulcers. *Annals of the New York Academy of Sciences*. 2018. 1411(1): 153.
37. Lipsky, B.A., Aragón-Sánchez, J., Diggle, M., Embil, J., Kono, S., Lavery, L., Senneville, É., Urbančič-Rovan, V., Van Asten, S., and Peters, E.J. IWGDF Guidance on the Diagnosis and Management of Foot Infections in Persons with Diabetes. *Diabetes/Metabolism Research and Reviews*. 2016. 32: 45-74.
38. Piaggese, A., Goretti, C., Iacopi, E., Clerici, G., Romagnoli, F., Toscanella, F., and Vermigli, C. Comparison of Removable and Irremovable Walking Boot to Total Contact Casting in Offloading the Neuropathic Diabetic Foot Ulceration. *Foot & Ankle International*. 2016. 37(8): 855-861.
39. Hajhosseini, B., Chiou, G.J., Dori, G., Fukaya, E., Chandra, V., Meyer, S., and Gurtner, G.C. Er: YAG Laser Versus Sharp Debridement in Management of Chronic Wounds: Effects on Pain and Bacterial Load. *Wound Repair and Regeneration*. 2020. 28(1): 118-125.
40. Chandler, L.A., Alvarez, O.M., Blume, P.A., Kim, P.J., Kirsner, R.S., Lantis, J.C., and Marston, W.A. Wound Conforming Matrix Containing Purified



- Homogenate of Dermal Collagen Promotes Healing of Diabetic Neuropathic Foot Ulcers: Comparative Analysis Versus Standard of Care. *Advances in Wound Care*. 2019. 9(2): 61-67.
41. Irawan, H., and Yasa, K.P. A Case Report of Diabetic Foot Ulcer Underwent an Autolytic Debridement Using Hydrogel and Hydrocellular Foam Combination. *Bali Med. J.* 2017. 6: S93-S96.
  42. Chen, C.Y., Wu, R.W., Hsu, M.C., Hsieh, C.J., and Chou, M.C. Adjunctive Hyperbaric Oxygen Therapy for Healing of Chronic Diabetic Foot Ulcers. *Journal of Wound, Ostomy and Continence Nursing*. 2017. 44(6): 536-545.
  43. Gottrup, F., Apelqvist, J., and Price, P. 'Outcomes in Controlled and Comparative Studies on Non-Healing Wounds: Recommendations. *Journal of Wound Care*. 2010. 19(6): 239.
  44. Vaupel, P., Kallinowski, F., and Okunieff, P. Blood Flow, Oxygen and Nutrient Supply, and Metabolic Microenvironment of Human Tumors: A Review. *Cancer Research*. 1989. 49(23): 6449-6465.
  45. Yu, G., Durduran, T., Lech, G., Zhou, C., Chance, B., Mohler, E.R., and Yodh, A.G. Time-Dependent Blood Flow and Oxygenation in Human Skeletal Muscles Measured with Noninvasive Near-Infrared Diffuse Optical Spectroscopies. *Journal of Biomedical Optics*. 2005. 10(2): 024027-02402712.
  46. Xia, J., Danielli, A., Liu, Y., Wang, L., Maslov, K., and Wang, L.V. Calibration-Free Quantification of Absolute Oxygen Saturation Based on the Dynamics of Photoacoustic Signals. *Optics Letters*. 2013. 38(15): 2800-2803.
  47. Huong, A.K.C. *Spectroscopic Analysis of Scattering Media Via Different Quantification Techniques*. Ph.D. Thesis. University of Nottingham; 2012.
  48. Cracowski, J.L., and Roustit, M. Current Methods to Assess Human Cutaneous Blood Flow: An Updated Focus on Laser-Based-Techniques', *Microcirculation*. 2016. 23(5): 337-344.
  49. Schreml, S., Szeimies, R., Prantl, L., Karrer, S., Landthaler, M., and Babilas, P. Oxygen in Acute and Chronic Wound Healing. *British Journal of Dermatology*. 2010. 163(2): 257-268.
  50. Demirtas, D., and Kucukosmanoglu, M. In Patients with Diabetic Foot, Improved Left Ventricular Functions are Detected by Strain

- Echocardiography After the Diabetic Foot Treatment: A Cross-Sectional Study. *Medicine*. 2019. 98(38).
51. Zhu, Y.Q., Wang, J., Tan, H.Q., Lu, H.T., Liu, F., Cheng, Y.S., Wei, L.M., Zhang, P.L., and Zhao, J.G. Runoff Detected by Magnetic Resonance Angiography as an Indicator for Better Recanalization Outcomes in Below-The-Knee Chronic Total Occlusions in Diabetic Patients. *Journal of Endovascular Therapy*. 2015. 22(2): 243-251.
  52. Arya, A.K., Tripathi, K., and Das, P. Promising Role of ANGPTL4 Gene in Diabetic Wound Healing. *The International Journal of Lower Extremity Wounds*. 2014. 13(1): 58-63.
  53. Liu, C., Van Netten, J.J., Van Baal, J.G., Bus, S.A., and Van Der Heijden, F. Automatic Detection of Diabetic Foot Complications with Infrared Thermography by Asymmetric Analysis. *Journal of Biomedical Optics*. 2015. 20 (2): 026003-026003.
  54. Fife, C.E., Horn, S.D., Smout, R.J., Barrett, R.S., and Thomson, B. A Predictive Model for Diabetic Foot Ulcer Outcome: The Wound Healing Index. *Advances In Wound Care*. 2016. 5(7) 279-287.
  55. Azuma, N. The Diagnostic Classification of Critical Limb Ischemia. *Annals of Vascular Diseases*. 2018. 11(4): 449-457.
  56. Wang, Z., Hasan, R., Firwana, B., Elraiyah, T., Tsapas, A., Prokop, L., Mills, J.L., and Murad, M.H. A Systematic Review and Meta-Analysis of Tests to Predict Wound Healing in Diabetic Foot. *Journal of Vascular Surgery*. 2016. 63(2): 29S-36S.
  57. Stewart, C., Frank, R., Forrester, K., Tulip, J., Lindsay, R., and Bray, R. A Comparison of Two Laser-Based Methods for Determination of Burn Scar Perfusion: Laser Doppler Versus Laser Speckle Imaging. *Burns*. 2005. 31 (6): 744-752.
  58. Nouvong, A., Hoogwerf, B., Mohler, E., Davis, B., Tajaddini, A., and Medenilla, E. Evaluation of Diabetic Foot Ulcer Healing with Hyperspectral Imaging of Oxyhemoglobin and Deoxyhemoglobin. *Diabetes Care*. 2009. 32(11): 2056-2061.
  59. Huong, A.K.C, and Ngu, X.T.I. In Situ Monitoring of Mean Blood Oxygen Saturation Using Extended Modified Lambert Beer Model. *Biomedical*

- Engineering: Applications, Basis and Communications*. 2015. 27(01): 1550004.
60. Sørensen, M.A., Petersen, L.J., Bundgaard, L., Toft, N., and Jacobsen, S. Regional Disturbances in Blood Flow and Metabolism in Equine Limb Wound Healing with Formation of Exuberant Granulation Tissue. *Wound Repair and Regeneration*. 2014. 22(5): 647-653.
  61. Jonsson, K., Jensen, J.A., Goodson 3rd, W., Scheuenstuhl, H., West, J., Hopf, H.W., and Hunt, T.K. Tissue Oxygenation, Anemia, and Perfusion in Relation to Wound Healing in Surgical Patients. *Annals of Surgery*. 1991. 214(5). 605.
  62. Meglinski, I.V., and Matcher, S.J. Quantitative Assessment of Skin Layers Absorption and Skin Reflectance Spectra Simulation in the Visible and Near-Infrared Spectral Regions. *Physiological Measurement*. 2002. 23(4): 741.
  63. Anderson, R.R., and Parrish, J.A. The Optics Of Human Skin. *Journal of Investigative Dermatology*. 1981. 77(1): 13-19.
  64. Barun, V., Ivanov, A., Volotovskaya, A., and Ulashchik, V. Absorption Spectra and Light Penetration Depth of Normal and Pathologically Altered Human Skin. *Journal of Applied Spectroscopy*. 2007. 74(3): 430-439.
  65. Tuchin, V. *Tissue Optics: Light Scattering Methods And Instruments For Medical Diagnosis*. 2nd Ed. Society of Photo-Optical Instrumentation Engineers (SPIE). 2007.
  66. Yusheng, F., Wang, Q., Yi, J., Song, D., and Xiang, X. A Numerical Model of Blood Flow Velocity Measurement Based on Finger Ring. *Journal of Healthcare Engineering*. 2018. 3916481-3916481.
  67. Wells, R.G., Marvin, B., Poirier, M., Renaud, J., and Ruddy, T.D. Optimization of SPECT Measurement of Myocardial Blood Flow with Corrections for Attenuation, Motion, and Blood Binding Compared with PET. *Journal of Nuclear Medicine*. 2017. 58(12): 2013-2019.
  68. Shung, K.K. *Diagnostic Ultrasound: Imaging and Blood Flow Measurements*. CRC Press. 2015.
  69. Goodman, J.W. Statistical Properties Of Laser Speckle Patterns. *Laser Speckle and Related Phenomena*. Springer. pp. 9-75; 1975.

70. Cheng, H., Luo, Q., Zeng, S., Chen, S., Cen, J., and Gong, H. Modified Laser Speckle Imaging Method with Improved Spatial Resolution. *Journal of Biomedical Optics*. 2003. 8(3): 559-565.
71. Fercher, A., and Briers, J.D. Flow Visualization by Means of Single-Exposure Speckle Photography. *Optics Communications*. 1981. 37(5): 326-330.
72. Parthasarathy, A.B., Tom, W.J., Gopal, A., Zhang, X., and Dunn, A.K. Robust Flow Measurement with Multi-Exposure Speckle Imaging. *Optics Express*. 2008. 16(3): 1975-1989.
73. Kazmi, S.S., Richards, L.M., Schrandt, C.J., Davis, M.A., and Dunn, A.K. Expanding Applications, Accuracy, and Interpretation of Laser Speckle Contrast Imaging of Cerebral Blood Flow. *Journal of Cerebral Blood Flow & Metabolism*. 2015. 35(7): 1076-1084.
74. Li, P., Ni, S., Zhang, L., Zeng, S., and Luo, Q. Imaging Cerebral Blood Flow through the Intact Rat Skull with Temporal Laser Speckle Imaging. *Optics Letters*. 2006. 31(12): 1824-1826.
75. Huang, Y.C., Ringold, T.L., Nelson, J.S., and Choi, B. Noninvasive Blood Flow Imaging for Real-Time Feedback During Laser Therapy of Port Wine Stain Birthmarks. *Lasers in Surgery and Medicine*. 2008. 40(3): 167-173.
76. Yang, B., Yang, O., Guzman, J., Nguyen, P., Crouzet, C., Osann, K.E., Kelly, K.M., Nelson, J.S., and Choi, B. Intraoperative, Real-Time Monitoring of Blood Flow Dynamics Associated with Laser Surgery of Port Wine Stain Birthmarks. *Lasers in Surgery and Medicine*. 2015. 47(6): 469-475.
77. Oberg, P.A. Laser-Doppler Flowmetry. *Critical Reviews in Biomedical Engineering*. 1989. 18(2): 125-163.
78. Shepherd, A. P. History Of Laser-Doppler Blood Flowmetry. In: Shepherd A.P and Oberg P.A. (Ed). *Laser-Doppler Blood Flowmetry*. Springer. pp. 1-16 ; 1990.
79. Fredriksson, I., Fors, C., and Johansson, J. *Laser Doppler Flowmetry-A Theoretical Framework*. Department of Biomedical Engineering, Linköping University. 2007. pp. 6-7.
80. Fredriksson, I., Larsson, M., and Strömberg, T. Measurement Depth and Volume in Laser Doppler Flowmetry. *Microvascular Research*. 2009. 78(1): 4-13.

81. Acharya, G., Mitra, A.K., and Cholkar, K. Nanosystems for Diagnostic Imaging, Biodetectors, and Biosensors. *Emerging Nanotechnologies for Diagnostics, Drug Delivery and Medical Devices*. Elsevier. 2017. pp. 217-248.
82. Bhang, H.E., Tsuchiya, N., Sysa-Shah, P., Winkelmann, C.T., and Gabrielson, K. In Vivo Small Animal Imaging: A Comparison with Gross and Histopathologic Observations in Animal Models. *Haschek and Rousseaux's Handbook of Toxicologic Pathology*. Elsevier. 2013. pp. 287-315.
83. Shah, J.V., and Shah, S. Upper Gastrointestinal Endoscopy in Early Diagnosis of Gastric Disorders. *International Journal of Contemporary Medical Research*. 2016. 3(7): 1943-1945.
84. Koo, H.J., Shin, J.H., Kim, H.J., Kim, J., Yoon, H.K., Ko, G.Y., and Gwon, D.I. Clinical Outcome of Transcatheter Arterial Embolization with N-Butyl-2-Cyanoacrylate for Control of Acute Gastrointestinal Tract Bleeding. *American Journal of Roentgenology*. 2015. 204(3): 662-668.
85. Tozzi, A., Bruni, A., and Bruni, C. *Endoscopic Guide, in Particular for Colonoscopy, and System for Endoscopy Comprising Such a Guide*. U.S. Patent Application 16/388, 471. 2019.
86. Kudo, S.E., Tamura, S., Nakajima, T., Yamano, H.O., Kusaka, H., and Watanabe, H. Diagnosis of Colorectal Tumorous Lesions by Magnifying Endoscopy. *Gastrointestinal Endoscopy*. 1996. 44(1): 8-14.
87. Gono, K. An Introduction to High-Resolution Endoscopy and Narrowband Imaging. *Comprehensive Atlas of High-Resolution Endoscopy and Narrowband Imaging*. 2017. 15: 7-15.
88. Turtaev, S., Leite, I.T., Altwegg-Boussac, T., Pakan, J.M., Rochefort, N.L., and Čižmár, T. High-Fidelity Multimode Fibre-Based Endoscopy for Deep Brain in Vivo Imaging. *Light: Science & Applications*. 2018. 7(1): 92.
89. Kobayashi, M., Ito, Y., Sakauchi, N., Oda, I., Konishi, I., and Tsunazawa, Y. Analysis of Nonlinear Relation for Skin Hemoglobin Imaging. *Optics Express*. 2001. 9(13): 802-812.
90. Gibson, A., Hebden, J., and Arridge, S.R. Recent Advances in Diffuse Optical Imaging. *Physics in Medicine & Biology*. 2005. 50(4): R1.

91. Sun, Y., Jiang, H., and O'Neill, B. Photoacoustic Imaging: An Emerging Optical Modality in Diagnostic and Theranostic Medicine. *J Biosens Bioelectron*. 2011. 2(108): 1000108-1000101.
92. Xia, J., Yao, J., and Wang, L.V. Photoacoustic Tomography: Principles And Advances. *Electromagnetic Waves*. Cambridge, Mass. 2014. 147: 1.
93. van den Berg, P.J. Integrated Photoacoustic/Ultrasound Imaging: Applications And New Techniques. Ph.D Thesis. University of Twente; 2017.
94. Laufer, J., Delpy, D., Elwell, C., and Beard, P. Quantitative Spatially Resolved Measurement of Tissue Chromophore Concentrations Using Photoacoustic Spectroscopy: Application to the Measurement of Blood Oxygenation and Haemoglobin Concentration. *Physics in Medicine and Biology*. 2006. 52(1): 141.
95. Larkin, P. *Infrared And Raman Spectroscopy: Principles And Spectral Interpretation*. Elsevier. 2017.
96. Butler, H.J., Ashton, L., Bird, B., Cinque, G., Curtis, K., Dorney, J., Esmonde-White, K., Fullwood, N.J., Gardner, B., and Martin-Hirsch, P.L. Using Raman Spectroscopy to Characterize Biological Materials. *Nature Protocols*. 2016. 11(4): 664.
97. Atkins, C.G., Buckley, K., Blades, M.W., and Turner, R.F. Raman Spectroscopy of Blood and Blood Components. *Applied Spectroscopy*. 2017. 71(5): 767-793.
98. Jermyn, M., Mok, K., Mercier, J., Desroches, J., Pichette, J., Saint-Arnaud, K., Bernstein, L., Guiot, M.-C., Petrecca, K., and Leblond, F. Intraoperative Brain Cancer Detection with Raman Spectroscopy in Humans. *Science Translational Medicine*. 2015. 7(274): 274ra219.
99. Pence, I., and Mahadevan-Jansen, A. Clinical Instrumentation and Applications of Raman Spectroscopy. *Chemical Society Reviews*. 2016. 45(7): 1958-1979.
100. Ugurbil, K.: What is Feasible with Imaging Human Brain Function and Connectivity Using Functional Magnetic Resonance Imaging. *Philosophical Transactions of the Royal Society B: Biological Sciences*. 2016. 371(1705): 20150361.



101. Saba, L., Porcu, M., Schmidt, B., and Flohr, T. Dual Energy CT: Basic Principles. In *Dual Energy CT in Oncology*. Springer. pp. 1-20; 2015.
102. Ross, A., and Bryskin, R. *Regional anesthesia, Smith's Anesthesia for Infants and Children*. Davis P.J., Cladis F.P., Motoyama E.K. (Ed). Philadelphia: Elsevier. pp. 403-444; 2011.
103. She, W., Cheung, T., Jenkins, C.R., and Irwin, M.G. Clinical Applications of High-Intensity Focused Ultrasound. *Hong Kong Medical Journal*. 2016. 22(4): 382-392.
104. Mace, E., Montaldo, G., Osmanski, B.-F., Cohen, I., Fink, M., and Tanter, M. Functional Ultrasound Imaging of the Brain: Theory and Basic Principles. *IEEE Transactions On Ultrasonics, Ferroelectrics, And Frequency Control*. 2013. 60(3): 492-506.
105. Deffieux, T., Demene, C., Pernot, M., and Tanter, M. Functional Ultrasound Neuroimaging: A Review of the Preclinical and Clinical State of the Art. *Current Opinion in Neurobiology*. 2018. 50: 128-135.
106. Pascual, T.N., Mercuri, M., El-Haj, N., Bom, H.H.-S., Lele, V., Al-Mallah, M.H., Luxenburg, O., Karthikeyan, G., Vitola, J., and Mahmarian, J.J. Nuclear Cardiology Practice in Asia: Analysis of Radiation Exposure and Best Practice for Myocardial Perfusion Imaging-Results from the IAEA Nuclear Cardiology Protocols Cross-Sectional Study (INCAPS). *Circulation Journal*. 2017. CJ-16-0677.
107. Berger, A. How Does It Work?: Positron Emission Tomography. *BMJ: British Medical Journal*. 2003. 326(7404): 1449.
108. Courchesne-Loyer, A., Croteau, E., Castellano, C.-A., St-Pierre, V., Hennebelle, M., and Cunnane, S.C. Inverse Relationship between Brain Glucose and Ketone Metabolism in Adults during Short-Term Moderate Dietary Ketosis: A Dual Tracer Quantitative Positron Emission Tomography Study. *Journal of Cerebral Blood Flow & Metabolism*. 2017. 37(7): 2485-2493.
109. Galldiks, N., Langen, K.-J., and Pope, W.B. From The Clinician's Point of View-What is The Status Quo of Positron Emission Tomography in Patients with Brain Tumors?. *Neuro-Oncology*. 2015. 17(11): 1434-1444.
110. Harada, R., Okamura, N., Furumoto, S., Yoshikawa, T., Arai, H., Yanai, K., and Kudo, Y. Use of a Benzimidazole Derivative BF-188 in Fluorescence

- Multispectral Imaging for Selective Visualization of Tau Protein Fibrils in The Alzheimer's Disease Brain. *Molecular Imaging and Biology*. 2014. 16(1): 19-27.
111. Wells, R.G., Timmins, R., Klein, R., Lockwood, J., Marvin, B., Wei, L., and Ruddy, T.D. Dynamic SPECT Measurement of Absolute Myocardial Blood Flow in a Porcine Model. *Journal of Nuclear Medicine*. 2014. 55(10): 1685-1691.
  112. Bailey, D.L., and Willowson, K.P. Quantitative SPECT/CT: SPECT Joins PET as a Quantitative Imaging Modality. *European Journal of Nuclear Medicine and Molecular Imaging*. 2014. 41(1): 17-25.
  113. Verberne, H.J., Acampa, W., Anagnostopoulos, C., Ballinger, J., Bengel, F., De Bondt, P., Buechel, R.R., Cuocolo, A., van Eck-Smit, B.L., and Flotats, A. EANM Procedural Guidelines for Radionuclide Myocardial Perfusion Imaging with SPECT And SPECT/CT: 2015 Revision. *European Journal of Nuclear Medicine and Molecular Imaging*. 2015. 42(12): 1929-1940.
  114. Zhang, Y., Liu, X., Wang, Q., Liu, D., Yang, C., Sun, J., and Rolfe, P. Influence of Extracerebral Layers on Estimates of Optical Properties with Continuous Wave Near Infrared Spectroscopy: Analysis Based On Multi-Layered Brain Tissue Architecture And Monte Carlo Simulation. *Computer Assisted Surgery*. 2019. pp. 1-7.
  115. Swartling, J., Dam, J.S., and Andersson-Engels, S. Comparison of Spatially and Temporally Resolved Diffuse-Reflectance Measurement Systems for Determination of Biomedical Optical Properties. *Applied Optics*. 2003. 42 (22): 4612-4620.
  116. Trebino, R. *Frequency-Resolved Optical Gating: The Measurement Of Ultrashort Laser Pulses*. Springer Science & Business Media. 2012.
  117. Saager, R.B., Baldado, M.L., Rowland, R.A., Kelly, K.M., and Durkin, A.J.: Method Using in Vivo Quantitative Spectroscopy to Guide Design and Optimization of Low-Cost, Compact Clinical Imaging Devices: Emulation and Evaluation of Multispectral Imaging Systems. *Journal Of Biomedical Optics*. 2018. 23(4): 046002.
  118. Vasefi, F., MacKinnon, N., and Farkas, D. Hyperspectral and Multispectral Imaging in Dermatology. *Imaging in Dermatology*. Elsevier. 2016. pp. 187-201.



119. Philimon, S.P. *An Alternative Means of Spectroscopic Imaging for Superficial Wound Healing Process Monitoring*. Master's Thesis. Universiti Tun Hussein Onn Malaysia; 2016.
120. Zhang, R., Verkruyse, W., Choi, B., Viator, J.A., Jung, B., Svaasand, L.O., Aguilar, G., and Nelson, J.S. Determination of Human Skin Optical Properties from Spectrophotometric Measurements Based on Optimization by Genetic Algorithms. *Journal of Biomedical Optics*. 2005. 10(2): 024030.
121. Pifferi, A., Taroni, P., Valentini, G., and Andersson-Engels, S. Real-Time Method for Fitting Time-Resolved Reflectance and Transmittance Measurements with a Monte Carlo Model. *Applied Optics*. 1998. 37(13): 2774-2780.
122. Swinehart, D. The Beer-Lambert Law. *Journal Of Chemical Education*. 1962. 39(7): 333.
123. Mäntele, W., and Deniz, E.: UV–VIS Absorption Spectroscopy: Lambert-Beer Reloaded. *Spectrochimica Acta. Part A, Molecular and Biomolecular Spectroscopy*. Elsevier. 2017. pp. 965-968.
124. Alabboud, I. *Human Retinal Oximetry Using Hyperspectral Imaging*. Ph.D. Thesis. Heriot-Watt University; 2009.
125. Pittman, R.N., and Duling, B.R. A New Method for the Measurement of Percent Oxyhemoglobin. *Journal of Applied Physiology*. 1975. 38(2): 315-320.
126. Huong, A., and Ngu, X. The Application of Extended Modified Lambert Beer Model for Measurement of Blood Carboxyhemoglobin and Oxyhemoglobin Saturation. *Journal of Innovative Optical Health Sciences*. 2014. 7(03): 1450026.
127. Chang, P., Walker, J., Hopcraft, K., Ablitt, B., and Jakeman, E. Polarization Discrimination for Active Imaging in Scattering Media. *Optics Communications*. 1999. 159(1): 1-6.
128. Zijlstra, W.G., Buursma, A., and van Assendelft, O.W. *Visible and Near Infrared Absorption Spectra of Human and Animal Haemoglobin: Determination and Application*. VSP. 2000.
129. Skoog, D.A., Holler, F.J., and Crouch, S.R. *Principles of Instrumental Analysis*. Cengage Learning. 2017.

130. Jagannath, R., Satija, A., Numa, N.M., Stockett, P.W., Joel, N., Lucht, R.P., and Bane, S.P. Application of a Streak Camera for Optical Emission Spectroscopy of Nanosecond Repetitively Pulsed Plasma Discharges. In *AIAA Scitech 2019 Forum*. 2019. pp. 0191.
131. Pershin, S., Grishin, M.Y., Lednev, V., Garnov, S., Bukin, V., Chizhov, P., Khodasevich, I., and Oshurko, V. Quantification of Distortion of the Water OH-Band Using Picosecond Raman Spectroscopy. *Laser Physics Letters*. 2018. 15(3): 035701.
132. Hirvonen, L.M., Becker, W., Milnes, J., Conneely, T., Smietana, S., Le Marois, A., Jagutzki, O., and Suhling, K. Picosecond Wide-Field Time-Correlated Single Photon Counting Fluorescence Microscopy with a Delay Line Anode Detector. *Applied Physics Letters*. 2016. 109(7): 071101.
133. Carboni, A., and Ferrero, A. A Fourier Transform-Based Frequency Estimation Algorithm. *IEEE Transactions on Instrumentation and Measurement*. 2018. 67(7): 1722-1728.
134. Seveck, E., Frisoli, J., Burch, C., and Lakowicz, J.R. Localization of Absorbers in Scattering Media by Use of Frequency-Domain Measurements of Time-Dependent Photon Migration. *Applied Optics*. 1994. 33(16): 3562-3570.
135. Tromberg, B.J., Svaasand, L.O., Tsay, T.-T., and Haskell, R.C. Properties of Photon Density Waves in Multiple-Scattering Media. *Applied Optics*. 1993. 32(4): 607-616.
136. Desmet, C.M., Lafosse, A., Vériter, S., Porporato, P.E., Sonveaux, P., Dufrane, D., Levêque, P., and Gallez, B. Application of Electron Paramagnetic Resonance (EPR) Oximetry to Monitor Oxygen in Wounds in Diabetic Models. *PloS One*. 2015. 10(12): e0144914.
137. Ning, B., Kennedy, M.J., Dixon, A.J., Sun, N., Cao, R., Soetikno, B.T., Chen, R., Zhou, Q., Shung, K.K., and Hossack, J.A. Simultaneous Photoacoustic Microscopy of Microvascular Anatomy, Oxygen Saturation, and Blood Flow. *Optics Letters*. 2015. 40(6): 910-913.
138. Yi, J., Liu, W., Chen, S., Backman, V., Sheibani, N., Sorenson, C.M., Fawzi, A.A., Linsenmeier, R.A., and Zhang, H.F. Visible Light Optical Coherence Tomography Measure Retinal Oxygen Metabolic Response to Systemic Oxygenation. *Light: Science & Applications*. 2015. 4(9): e334-e334.

139. Yazdi, H.S., O'Sullivan, T.D., Leproux, A., Hill, B., Durkin, A., Telep, S., Lam, J., Yazdi, S.S., Police, A.M., and Carroll, R.M. Mapping Breast Cancer Blood Flow Index, Composition, and Metabolism in a Human Subject Using Combined Diffuse Optical Spectroscopic Imaging and Diffuse Correlation Spectroscopy. *Journal Of Biomedical Optics*. 2017. 22(4): 045003.
140. Fondi, K., Wozniak, P.A., Howorka, K., Bata, A.M., Aschinger, G.C., Popa-Cherecheanu, A., Witkowska, K.J., Hommer, A., Schmidl, D., and Werkmeister, R.M. Retinal Oxygen Extraction in Individuals with Type 1 Diabetes with No or Mild Diabetic Retinopathy. *Diabetologia*. 2017. 60(8): 1534-1540.
141. Felder, A.E., Wanek, J., Tan, M.R., Blair, N.P., and Shahidi, M. A Method for Combined Retinal Vascular and Tissue Oxygen Tension Imaging. *Scientific Reports*. 2017. 7(1): 10622.
142. Merčep, E., Deán-Ben, X.L., and Razansky, D. Imaging of Blood Flow and Oxygen State with a Multi-Segment Optoacoustic Ultrasound Array. *Photoacoustics*. 2018. 10: 48-53.
143. Ghijsen, M., Lentsch, G.R., Gioux, S., Brenner, M., Durkin, A.J., Choi, B., and Tromberg, B.J. Quantitative Real-Time Optical Imaging of the Tissue Metabolic Rate of Oxygen Consumption. *Journal of Biomedical Optics*. 2018. 23(3): 036013.
144. Lucero, A.A., Addae, G., Lawrence, W., Neway, B., Credeur, D.P., Faulkner, J., Rowlands, D., and Stoner, L. Reliability of Muscle Blood Flow and Oxygen Consumption Response from Exercise Using Near-Infrared Spectroscopy. *Experimental Physiology*. 2018. 103(1): 90-100.
145. Saidian, M., Lakey, J.R., Ponticorvo, A., Rowland, R., Baldado, M., Williams, J., Pronda, M., Alexander, M., Flores, A., and Shiri, L. Characterisation of Impaired Wound Healing in a Preclinical Model of Induced Diabetes Using Wide-Field Imaging and Conventional Immunohistochemistry Assays. *International Wound Journal*. 2019. 16(1): 144-152.
146. Lin, B.S., Chang, C.C., Tseng, Y.H., Li, J.R., Peng, Y.S., and Huang, Y.K. Using Wireless Near-Infrared Spectroscopy to Predict Wound Prognosis in Diabetic Foot Ulcers. *Advances in Skin & Wound Care*. 2020. 33(1): 1-12.

147. Basak, K., Dey, G., Mahadevappa, M., Mandal, M., Sheet, D., and Dutta, P.K. Learning of Speckle Statistics for in Vivo and Noninvasive Characterization of Cutaneous Wound Regions Using Laser Speckle Contrast Imaging. *Microvascular Research*. 2016. 107: 6-16.
148. Rege, A., Thakor, N.V., Rhie, K., and Pathak, A.P.: 'In Vivo Laser Speckle Imaging Reveals Microvascular Remodeling and Hemodynamic Changes During Wound Healing Angiogenesis. *Angiogenesis*. 2012. 15(1) 87-98.
149. Wang, L., Cull, G.A., Piper, C., Burgoyne, C.F., and Fortune, B. Anterior and Posterior Optic Nerve Head Blood Flow in Nonhuman Primate Experimental Glaucoma Model Measured by Laser Speckle Imaging Technique and Microsphere Method Optic Nerve Blood Flow in Experimental Glaucoma. *Investigative Ophthalmology & Visual Science*. 2012. 53(13): 8303-8309.
150. Shiba, C., Shiba, T., Takahashi, M., Matsumoto, T., and Hori, Y. Relationship Between Glycosylated Hemoglobin A1c and Ocular Circulation by Laser Speckle Flowgraphy in Patients with/without Diabetes Mellitus. *Graefe's Archive for Clinical and Experimental Ophthalmology*. 2016. 254(9): 1801-1809.
151. He, H., Tang, Y., Zhou, F., Wang, J., Luo, Q., and Li, P. Lateral Laser Speckle Contrast Analysis Combined with Line Beam Scanning Illumination to Improve the Sampling Depth of Blood Flow Imaging. *Optics Letters*. 2012. 37 (18): 3774-3776.
152. Cooper, R., Selb, J., Gagnon, L., Phillip, D., Schytz, H.W., Iversen, H.K., Ashina, M., and Boas, D.A. A Systematic Comparison of Motion Artifact Correction Techniques for Functional Near-Infrared Spectroscopy. *Frontiers in Neuroscience*. 2012. 6: 147.
153. Sdobnov, A., Bykov, A., Popov, A., Zhrebtssov, E., and Meglinski, I. Investigation of Speckle Pattern Dynamics By Laser Speckle Contrast Imaging. In *Biophotonics: Photonic Solutions for Better Health Care VI*. International Society for Optics and Photonics. 2018. vol. 10685, pp. 1068509.
154. Lagarias, J.C., Reeds, J.A., Wright, M.H., and Wright, P.E. Convergence Properties of the Nelder-Mead Simplex Method in Low Dimensions. *SIAM Journal on Optimization*. 1998. 9(1): 112-147.

155. Dutta, S.D., and Maria, R. Pulse Oximetry. A New Tool in Pulpal Vitality Testing. *People's Journal of Scientific Research*. 2013. 6: 49-52.
156. Philimon, S.P., Huong, A.K.C, and Ngu, X.T.I. Multispectral Imaging System for Quantitative Assessment of Transcutaneous Blood Oxygen Saturation. *Jurnal Teknologi*. 2015. 77(7): 37-41.
157. Philimon, S.P, Huong, A.K.C., and Ngu, X.T.I. An Alternative Wavelength Range for Noninvasive Assessment of Wound Tissue Oxygenation Status. *International Journal of Engineering and Technology, Science Publishing Corporation*. 2018. 7(4.26): 73-77.
158. Huong, A., and Ngu, X. Neural Network Approach For Rapid Prediction of Transcutaneous Oxygen Saturation. In *2019 IEEE 9<sup>th</sup> Symposium on Computer Applications & Industrial Electronics (ISCAIE)*. IEEE. 2019. pp. 239-243.
159. Aikio, M. *Hyperspectral Prism-Grating-Prism Imaging Spectrograph*. Ph.D Thesis. University of Oulu; 2001.
160. Huong, A., Philimon, S., and Ngu, X. Noninvasive Monitoring of Temporal Variation in Transcutaneous Oxygen Saturation for Clinical Assessment of Skin Microcirculatory Activity. In *International Conference for Innovation in Biomedical Engineering and Life Sciences*. Springer, Singapore, 2015. pp. 248-251.
161. Uemura, N., Okazaki, M., and Mori, H. Anatomical and Histological Study to Determine the Border of Sole Skin. *Surgical and Radiologic Anatomy*. 2016. 38 (7): 767-773.
162. Lu, W.C., Lu, S.H., Chen, M.F., Fu, T.C., Lin, K.P., and Tsai, C.L. Portable Near-Infrared Spectroscopy for Detecting Peripheral Arterial Occlusion. *Precision Medicine Powered by pHealth and Connected Health*. Springer. 2018. pp. 109-113.
163. Brown, A.D., McMorris, C.A., Longman, R.S., Leigh, R., Hill, M.D., Friedenreich, C.M., and Poulin, M.J. Effects of Cardiorespiratory Fitness and Cerebral Blood Flow on Cognitive Outcomes in Older Women. *Neurobiology of Aging*. 2010. 31(12): 2047-2057.
164. Kirkpatrick, S.J., Duncan, D.D., and Wells-Gray, E.M. Detrimental Effects of Speckle-Pixel Size Matching in Laser Speckle Contrast Imaging. *Optics Letters*. 2008. 33(24): 2886-2888.



165. Molnár, E., Molnár, B., Lohinai, Z., Tóth, Z., Benyó, Z., Hricisák, L., Windisch, P., and Vág, J. Evaluation of Laser Speckle Contrast Imaging for the Assessment of Oral Mucosal Blood Flow Following Periodontal Plastic Surgery: An Exploratory Study. *BioMed Research International*. 2017. pp. 4042902-4042902.
166. Forrester, K.R., Tulip, J., Leonard, C., Stewart, C., and Bray, R.C. A Laser Speckle Imaging Technique for Measuring Tissue Perfusion. *IEEE Transactions on Biomedical Engineering*. 2004. 51(11): 2074-2084.
167. Briers, J.D. Laser Doppler, Speckle and Related Techniques for Blood Perfusion Mapping and Imaging. *Physiological Measurement*. 2001. 22(4): R35.
168. Tom, W.J., Ponticorvo, A., and Dunn, A.K. Efficient Processing of Laser Speckle Contrast Images. *IEEE Transactions on Medical Imaging*. 2008. 27(12): 1728-1738.
169. Hecht, J. Helium Neon Lasers Flourish in Face of Diode-Laser Competition. *Laser Focus World*. 1992. 28(11): 99-108.
170. Louie, A., Feiner, J.R., Bickler, P.E., Rhodes, L., Bernstein, M., and Lucero, J. Four Types of Pulse Oximeters Accurately Detect Hypoxia During Low Perfusion and Motion. *Anesthesiology: The Journal of the American Society of Anesthesiologists*. 2018. 128(3): 520-530.
171. Philimon, S.P., Huong, A.K., and Ngu, X.T. Investigation of Multispectral Imaging Technique for Optical Monitoring of Mean Blood Oxygen Saturation. *ARNP Journal of Engineering and Applied Sciences*. Asian Research Publishing Network. 2016. 11(6): 3951-3956.
172. Vogel, A., Chernomordik, V.V., Demos, S.G., Pursley, R., Little, R.F., Tao, Y., Gandjbakhche, A.H., Yarchoan, R., Riley, J.D., and Hassan, M. Using Noninvasive Multispectral Imaging to Quantitatively Assess Tissue Vasculature. *Journal of Biomedical Optics*. 2007. 12(5): 051604-051613.
173. Ciarlillo, D., Celeste, C., Carmeliet, P., Boerboom, D., and Theoret, C. A Hypoxia Response Element in the Vegfa Promoter is Required for Basal Vegfa Expression in Skin and for Optimal Granulation Tissue Formation During Wound Healing in Mice. *PloS One*. 2017. 12(7): e0180586.
174. Boike, A., Maier, M., and Logan, D. *Prevention and Treatment of Leg and Foot Ulcers in Diabetes Mellitus*. Current Clinical Medicine. 2010.

175. Sen, C.K. Wound Healing Essentials: Let There Be Oxygen. *Wound Repair and Regeneration*. 2009. 17(1): 1-18.
176. Kimmel, H.M., Grant, A., and Ditata, J. The Presence of Oxygen in Wound Healing. *Wounds: A Compendium of Clinical Research and Practice*. 2016. 28(8): 264-270.
177. Chawla, A., Chawla, R., and Jaggi, S. Microvascular and Macrovascular Complications in Diabetes Mellitus: Distinct Or Continuum?. *Indian Journal of Endocrinology and Metabolism*. 2016. 20(4): 546.
178. Witte, D., Tesfaye, S., Chaturvedi, N., Eaton, S., Kempler, P., Fuller, J., and Group, E.P.C.S. Risk Factors for Cardiac Autonomic Neuropathy in Type 1 Diabetes Mellitus. *Diabetologia*. 2005. 48(1): 164-171.
179. Andersen, S.T., Witte, D.R., Fleischer, J., Andersen, H., Lauritzen, T., Jørgensen, M.E., Jensen, T.S., Pop-Busui, R., and Charles, M. Risk Factors for the Presence and Progression of Cardiovascular Autonomic Neuropathy in Type 2 Diabetes: ADDITION-Denmark. *Diabetes Care*. 2018. 41(12): 2586-2594.
180. Roustit, M., Loader, J., Baltzis, D., Zhao, W., and Veves, A. Microvascular Changes in the Diabetic Foot. *The Diabetic Foot*. Springer. 2018. pp. 173-188.
181. Mustoe, T. Understanding Chronic Wounds: A Unifying Hypothesis on Their Pathogenesis and Implications for Therapy. *The American Journal of Surgery*. 2004. 187(5): S65-S70.
182. Rendell, M.S., Milliken, B.K., Finnegan, M.F., Finney, D.A., and Healy, J.C. The Skin Blood Flow Response in Wound Healing. *Microvascular Research*. 1997. 53(3): 222-234.
183. Wigington, G., Ngo, B., and Rendell, M. Skin Blood Flow in Diabetic Dermopathy. *Archives of Dermatology*. 2004. 140(10): 1248-1250.
184. Isakson, M., De Blacam, C., Whelan, D., McArdle, A., and Clover, A. Mesenchymal Stem Cells and Cutaneous Wound Healing: Current Evidence and Future Potential. *Stem Cells International*. 2015.